

Table of Contents

Introductory Information	
Schedule	ii
Instructor Contact Information	iii
Evaluation and Assessment	iv
Laboratory Report Guidelines and Format	vi
Course Policies	vii
University Policy on Academic Integrity	viii
Safety Regulations	ix
Fire and Earthquake Evacuation Procedures	xi
Laboratory Procedures	
Lab 1 – Isolation and Identification of Peptides and Proteins	1-1
Lab 2 – Chromatin Immunoprecipitation (ChIP) Analysis of YEF3	2-1
Appendix 2 – Glossary of Terms	2-26
Lab 3 – Immunological Characterization of Breast Cancer Cell Lines	3-1
Appendix 3 – Glossary of Terms	3-31
Equipment Usage and Instructions	
Appendix A – Fumehood Guidelines	A-1
Appendix B – Laminar Flow Hoods and Biological Safety Cabinets	B-1
Appendix C – Working in a Biological Safety Cabinet	C-1
Appendix D – Gen5 & KCJunior Software Instructions	D-1

BCMB 406A Laboratory Schedule, Summer 2017

Week	Dates	Lab(s)	Day 1 (5 hours)	Day 2 (2 hours)	Due Dates
1	May 8-9	Introduction and Safety talk Lab 1: Isolation and Identification of Peptides and Proteins	Bradford assay	1 st dimension of 2D-PAGE	
	May 10-11	Lab 1: Isolation and Identification of Peptides and Proteins	2 nd dimension of 2D-PAGE and HPLC	Gel imaging & spot excision	
2	May 15-16	Lab 1: Isolation and Identification of Peptides and Proteins	Tryptic digestion and HPLC	Zip-tip plate spotting	
	May 17-18	Lab 1: Isolation and Identification of Peptides and Proteins Lab 2: Chromatin Immunoprecipitation Analysis of YEF3	Lab 1: Mass spectrometry Lab 2: Chromatin prep, MNase digestion and reverse cross-linking	Lab 1: Protein identification	
3	May 22-23	Lab 2: Chromatin Immunoprecipitation Analysis of YEF3	Victoria Day (no labs)	Lab 2: DNA purification and agarose gel	Day 2: Lab 1 report
	May 24-25	Lab 2: Chromatin Immunoprecipitation Analysis of YEF3	Immunoprecipitation (IP)	IP and reverse cross-linking	Day 1: Lab 2 Prelab assign.
4	May 29-30	Lab 2: Chromatin Immunoprecipitation Analysis of YEF3	Lab 1 Study Block	Lab 1 Midterm	
	May 31 - Jun 1	Lab 2: Chromatin Immunoprecipitation Analysis of YEF3 Lab 3: Immunological Characterization of Cancer Cell Lines	Lab 2: DNA purificat'n, QPCR and PCR Lab 3: Prelab talk	Agarose gel and QPCR data analysis	Day 1: Lab 3 calculation problem set
5	June 5-6	Lab 3: Immunological Characterization of Cancer Cell Lines	Cell culture and cell staining	Finish cell staining	Day 2: Lab 2 report
	June 7-8	Lab 3: Immunological Characterization of Cancer Cell Lines	Cell culture and T cell assay set-up	T cell assay development	
6	June 12-13		Lab 2 Study Block	Lab 2 Midterm	
	June 14-15	Lab 3: Immunological Characterization of Cancer Cell Lines	FlowJo Tutorial and data analysis	Data Analysis	Day 1: FlowJo exercise
7	June 19-20				Day 2: Lab 3 report
	June 21-22				
8	June 26-27		Lab 3 Study Block	Lab 3 Midterm	

Instructor Contact Information

Lab	Instructor	Email	Phone	Office
1	Glen Pryhitka	pryhitka@uvic.ca	250-721-7076	Petch 179a
2	Val Kerr	valk@uvic.ca	250-472-5119	Petch 179b
3	Erika Wall	ewall@uvic.ca	250-472-5119	Petch 179b

Each instructor is responsible for a different lab as indicated above. Please make sure that you address any concerns or questions to the appropriate instructor.

CourseSpaces

<http://coursespaces.uvic.ca/my/>

Enter: NetLink-ID and Password

Select: 201705 BCMB 406A B01

Important information for the course and each lab is posted on CourseSpaces. Announcements are frequently sent to students, so please make sure that you check the email you have on record.

Departmental Web Page

<http://web.uvic.ca/biochem>

Occupational Health and Safety

250-721-8971

<http://ohs.uvic.ca>

Police, Fire, Ambulance 911

Campus Security

250-721-7599

<http://web.uvic.ca/security>

University Health Services

250-721-8492

<http://www.uvic.ca/services/health>

Evaluation and Assessment

Percentage Breakdown for the Course:

Lab Reports	20 %	see pages v and vi for more details
Exams	50 %	see page v for more details
Practical Assessment	20 %	see page v for more details
Laboratory Journal	10 %	see page v for more details

Final Course Percentages:

A final percentage will be calculated for the course based on the above criteria. All percentages will be rounded to the nearest whole number. For example, a calculated percentage of 79.49% will be recorded as 79% whereas 79.50% will be recorded as 80%.

Letter Grades will be assigned as follows:

90 - 100	A+
85 - 89	A
80 - 84	A-
77 - 79	B+
73 - 76	B
70 - 72	B-
65 - 69	C+
60 - 64	C
50 - 59	D
0 - 49	F (or N*)

* N grades

Students who have completed the following course requirements will be considered to have completed the course and will be assigned a final percentage and letter grade.

- *In class lab work (all Day 1 and Day 2 components of the practical work must be performed)*
- *Midterm exam*
- *Final exam*

Failure to complete one or more of these elements will result in a grade of "N" regardless of the cumulative percentage on other elements of the course. An N is a failing grade, and it factors into a student's GPA as 0. The maximum percentage that can accompany an N on a student's transcript is 49.

Lab Reports (20%): Marks for the lab reports will be assigned as indicated below.

Laboratory Report	Marks
Lab 1 – Isolation and Identification of Proteins and Peptides	30
Lab 2 – Chromatin Immunoprecipitation (ChIP) Analysis	30
Lab 3 – Immunological Characterization of Breast Cancer Cell Lines	40
Total Marks	100

Lab Exams (50%):

There will be three midterms in this course. Each midterm will be two hours in length and will only cover material from one lab. Midterm 1 and 2 will be worth 15% each and Midterm 3 will be worth 20%.

Midterm	Date	Lab covered	Percent
1	Tues. May 30	Lab 1	15 %
2	Tues. June 13	Lab 2	15 %
3	Tues. June 27	Lab 3	20 %

Practical Evaluation (20%):

Practical assessments will be done for each student by laboratory instructors and teaching assistants and will consist of:

- Frequent pre-lab quizzes to assess preparedness
- Pre-lab and in-lab assignments, calculations and problem sets
- Frequent evaluation of experimental results to assess technique
- Thoroughness of clean-up at each bench after the lab

Weighting of these assignments will vary based on the discretion of the instructors.

Maintenance of a Laboratory Journal (10%):

You are expected to provide a hard cover or spiral bound notebook to be used as a laboratory journal. This book is dedicated to recording both raw data generated while performing the labs and processed data used to create figures and tables. Your lab journal must be brought to every lab session. Data for each lab will be marked during the midterm exam.

Please write in ink and include all relevant information, such as:

- Date and title of the experiment
- Unknown numbers
- Pre-lab or in-lab calculations
- Detailed procedural steps used when not working directly from the lab manual
- All raw data you (and/or your partner) generate with important information included
- Experimental conditions (temperature, time, wavelengths, etc...)
- Loading order and volumes of samples put onto gels, etc...
- Changes to the procedure or mistakes/errors made
- Handouts of data and/or copies of student data posted on CourseSpaces
- **Observations and interpretations of results (this is new compared to expectations in second and third year lab courses in this department)**

Note: Portions of marked lab reports will not be considered as lab journal entries.

Laboratory Report Guidelines and Format

- **All laboratory reports must be written up individually.**
- Text should be typed and double-spaced, with margins no smaller than 1.9 cm (0.75")
- Use 12 point font of a standard style such as Arial or Times New Roman.
- All written text should be concise, well written and proofread for grammar and spelling.

Below is a general outline of what should be included in the following portions of a lab report in this course. Each lab report will have a different set of requirements so be sure to read the instructions for each submission carefully. Refer to CourseSpaces for a more detailed description of lab report expectations.

- Abstract:** State the purpose of the experiment(s) and put it in context. Summarize the methods, results and conclusions of the research.
- Introduction:** In a few well written paragraphs, state the purpose and introduce the main concepts of the lab by defining important terms and explaining new ideas. As well, briefly describe and indicate the purpose of performing the individual techniques or experiments.
- Methods:** Refer to the source of the procedure in proper citation format (most of the time this will be the lab manual). For procedures that you have designed, briefly summarize techniques and materials used so that someone could repeat the experiment (minor details are not necessary). Include any *significant* modifications that were communicated to you either verbally or in written form. Also, include mistakes that were made by you, your partner or other individuals that may have affected your results.
- Results:** Organize data in the form of fully labeled tables, graphs or figures. State the results in written form in a *Results Summary* placed before the figures, drawing attention to the key results. Reserve *all* interpretation for the discussion. Tables, graphs and figures should be numbered in the order in which they are cited in the text.
- Discussion:** The discussion should provide an interpretation of your results. Be concise. Keep discussion relevant to the data generated from your own experiment (and that of your classmates when appropriate). Incorporate into your discussion the answers to any relevant questions that appear in the lab manual. In a couple of sentences, draw a conclusion based on the results of the experiment.
- Appendix:** Include raw data, calculations and other information that is relevant. Remember to number your appendices and include a basic title for each.
- References:** Online links to all relevant reference papers can be found on CourseSpaces. Laboratory reports must be referenced following the format of the *Journal of Molecular Biology*. This is the same format we use in the lab manual.

Please note: A lab report submission consists of both a hard copy and electronic copy. Hard copies of reports are to be submitted to the instructor, whereas electronic copies are to be uploaded to CourseSpaces. Both must be submitted by **12:30 pm** on the designated due date, otherwise late penalties will apply.

Course Policies

Attendance

Laboratory attendance is compulsory. Failure to attend a lab without a written medical excuse or proof of undue hardship will result in a mark of '**N**' (**incomplete**) for the course. If an absence is granted by the instructor, it is the student's responsibility to make arrangements with the lab instructor **prior** to the lab period to make up the missed lab work. Excused students are also responsible for maintaining their lab journal and obtaining the data needed to write up the lab report.

It is important to arrive on time. Students who arrive after a pre-lab quiz has begun will not be given extra time to complete the quiz. No makeup quizzes will be given for students who arrive after a quiz is over. The instructor reserves the right to refuse late arrivals or withhold practical marks associated with that lab if a student shows up late without a legitimate reason.

Late Assignments or Lab Reports

Late assignment or lab reports (either the hard copy or the electronic copy) will be penalized 10% per day and will not be accepted after one week (7 days) following the designated due date. Late lab reports or assignments can be emailed to the instructor on days when the university is closed but a hard copy must be submitted by noon on the first working day after the emailed submission.

Re-marking Policy

Request to have assignments (lab reports, quizzes, exams, journal entries, practical assessments, etc.) reviewed/remarked must be made within one week of the assignments being made available. Students are expected to promptly pick up assignments after marking is completed, either in lab or from the instructor. Any resubmissions will involve remarking of the entire assignment and students will be given the grade assigned after the second review.

Accessibility Statement

Students with diverse learning styles and needs are welcome in this course. If you have a disability/health consideration that may require accommodations, please feel free to approach an instructor and/or the Resource Centre for Students with a Disability (<http://rcsd.uvic.ca/>) as soon as possible. The RCSD staff is available by appointment to assess specific needs, provide referrals and arrange appropriate accommodations. The sooner you let us know your needs the quicker we can assist you in achieving your learning goals in this course.

Department Policies

This course abides by the departmental policies outlined in the document "Departmental Information and Policies" located on CourseSpaces.

Course Experience Survey (CES)

We value your feedback. Towards the end of term, you will have the opportunity to complete an anonymous survey regarding your learning experience. The survey provides vital feedback regarding the course and our teaching, as well as helping the department improve the overall program for future students. The survey is accessed via MyPage and can be done on your laptop, tablet, or mobile device. We will provide more detailed information nearer the time but ask that you think about this important activity during the course.

University Policy on Academic Integrity

Suspected cases of plagiarism or cheating will be documented and submitted to the department chair for penalty assessment as described in the UVic calendar (2015-2016).

Plagiarism

A student commits plagiarism when he or she:

- submits the work of another person as original work
- gives inadequate attribution to an author or creator whose work is incorporated into the student's work, including failing to indicate clearly the inclusion of another individual's work
- paraphrases material from a source without sufficient acknowledgement as described above

Falsifying Materials Subject to Academic Evaluation

Falsifying materials subject to academic evaluation includes, but is not limited to:

- fraudulently manipulating laboratory processes, electronic data or research data in order to achieve desired results
- using work prepared by someone else and submitting it as one's own
- citing a source from which material was not obtained
- using a quoted reference from a non-original source while implying reference to the original source
- submitting false records, information or data, in writing or orally

Cheating on Assignments, Tests and Examinations

Cheating includes, but is not limited to:

- copying the answers or other work of another person
- sharing information or answers when doing take-home assignments, tests and examinations except where the instructor has authorized collaborative work
- having in an examination or test any materials or equipment other than those authorized by the examiners

Collaborative Work (Penalties) * See below for details specific to this course.

In cases in which an instructor has provided clear written instructions prohibiting certain kinds of collaboration on group projects, instances of prohibited collaboration on a substantial part of the assignment should result in a grade of zero for the assignment, while instances of prohibited collaboration on the bulk of the assignment should result in a grade of F for the course.

*** In BCMB 406A, it is required that all assignments and lab reports be written up independently. Collaborative work is only allowed in specific situations when students are directed to share data by the instructor. Students can discuss thoughts and ideas with other students but all written work must be original. Be sure to submit work that is entirely your own.**

Safety Regulations

Work in a microbiology laboratory involves exposure to living microorganisms, many of which must be considered as potential pathogens. Personal recognition of safety and the acceptance of certain precautions are therefore necessary prerequisites to working in the laboratory.

1. **Access to the laboratory is limited to instructors and students.**
2. **No eating, drinking or smoking in the laboratory.** Keep paper, pencils, fingers, and other objects out of the mouth.
3. **Safety glasses must be worn at all times.**
4. **Laboratory coats must be worn and properly fastened by all personnel working in the laboratory and must not be worn in public places.** Laboratory coats will be provided for you and will be shared between sections. If you would prefer a lab coat of your own, you will need to bring one to your first lab section. It will be kept in the lab until the end of the course, at which time it will be autoclaved and available for pick-up.
5. **Open-toed shoes or sandals should not be worn and bare legs are not allowed.** Capri pants, skirts and shorts are only allowed if they cover the knees when you are sitting down.
6. **Lab doors and windows must be closed when working with Level 2 organisms.**
7. **Personal items (coats, bags, etc...) must be kept away from the work area.**
8. **Wash hands before and after completing all lab work.**
9. **Long hair must be tied back** to protect against burning and falling into stains, chemicals or bacterial cultures.
10. **Gloves must be worn in the following situations:**
 - Working with Level 2 organisms
 - Working with potentially harmful reagents
 - If you have open cuts or abrasions on your hands
11. **Gloves and lab coats are not to be worn outside of the lab.**
12. **Remove gloves when working with communal equipment and computers.**
13. **Bunsen burners are to remain off when not in use.**
14. **Dispose of all liquids into the designated waste containers.** No liquids are to be poured down the sink (except water).
15. **Contaminated liquid waste must be autoclaved prior to disposal.**

16. **Dispose of infectious solid waste in the yellow biohazard buckets for autoclaving.**
This includes pipette tips, agar plates, contaminated gloves or paper towels, etc...

17. **Report any accidents or safety concerns to an instructor immediately.**

- If skin comes into contact with chemicals, wash immediately with cold running water for at least 10 min.
- In the event of a bacterial spill, pour an equal volume of bench disinfectant on top of the spill and allow it to sit for five minutes. Clean up the spill wearing gloves and using a no-touch technique. Discard all waste in a yellow biohazard bucket for autoclaving. **WASH YOUR HANDS with hand disinfectant and soap.**
- Do not pick up broken glass. The instructor will do this.
- If something has splashed in your eyes, rinse them at the eye wash station for at least 20 min.

18. **Mouth pipetting is prohibited.** A safety bulb or pipettor must be used.

19. **Sitting on laboratory work surfaces is prohibited.**

20. **Use of cellular devices is not allowed in the lab.**

21. **Note the location of the following safety equipment:**

- Eye wash station
- Safety shower
- Fire extinguisher
- Telephone
- Fire alarm

22. **Before leaving the laboratory:**

- Place all cultures and other contaminated materials to be discarded in the appropriate containers for sterilization in the autoclave
- Put your experimental materials (**labeled!**) in the appropriate bins or racks for incubation or storage
- Place contents of “tip discard” and used microfuge tubes into yellow biohazard bucket
- Rinse all glassware and place in the appropriate bin(s)
- Check that gas, water and microscope lights are turned off
- Wash the bench top with bench disinfectant
- Wash your hands thoroughly with hand disinfectant and/or soap

Experiments conducted in a microbiology laboratory involve the handling of pathogenic organisms. Failure to handle and dispose of these organisms correctly may lead to infection, injury or even serious illness. For the safety of everyone, it is required that you understand and follow the appropriate laboratory procedures as outlined by your laboratory instructor.

Your signature on the Biochemistry and Microbiology Lab Safety Form is your acknowledgement that you have read the safety regulations and agree to abide by them.

Fire Evacuation Procedure

If you discover a fire:

- Activate the nearest fire alarm pull station.
- Call **911** and Campus Security Services at **7599**. State your name and location.
- Evacuate the building.

If you hear a fire alarm:

- If possible secure equipment and close windows and doors.
- Follow the established evacuation route. Do not use elevators.
- Meet at your designated Emergency Evacuation Site.
- Do not re-enter the building until permission is given by the Fire Department.

If you cannot evacuate:

- Close the doors between you and the fire.
- If possible call **911** and advise the Fire Department of your situation.
- Hang clothing or a cloth from a window to alert emergency response personnel.

Earthquake Evacuation Procedure

During an Earthquake:

- Get away from windows and heavy objects.
- Duck, cover and hold on. Crouch low to the ground; protect head with your arms; seek cover under and hold onto heavy furniture. Watch for moving objects.
- If you are in an interior hallway, stay there and crouch against the wall. Watch for swinging doors.

After an Earthquake:

- After the shaking stops wait 60 seconds then evacuate the building. Do not use elevators.
- Make your way to the UVic Campus Assembly Area to receive instructions and information. Keep away from power lines and buildings to avoid falling debris.
- Report any injuries to Campus Security Services.